Fig. 3

SDS-PAGE Analysis of
2H7 scFvIgG1 (SSS-S)H WCH2 WCH3 Protein.

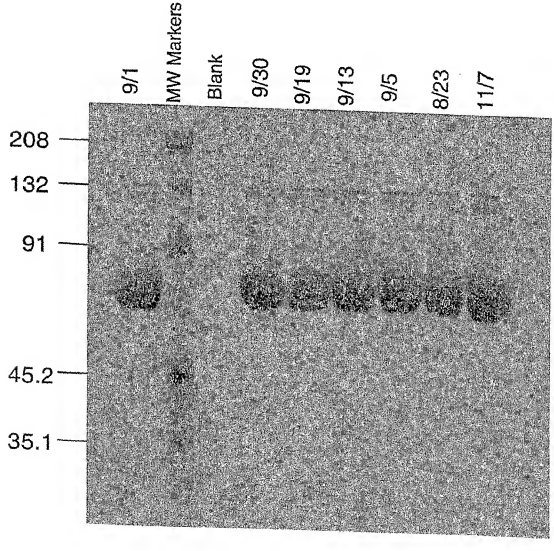


Fig. 4A

Complement Mediated B Cell Killing After Binding of CD20-targeted 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3:

2H7scFv-Ig Concentration	RAMOS # live cells/total cells		BJAB # live cells/total cells	
20 μg/ml + complement	-	0.16	-	0.07
5 μg/ml + complement	-	0.2	-	N.D.
1.25 μg/ml + complement	-	0.32	-	0.1
Complement alone	-	0.98	-	0.94

*Viability was determined by trypan blue exclusion and is tabulated as the fraction of viable cells out of the total number of cells counted.

**N.D. (not determined).

Fig. 4B

Antibody-dependent cellular cytotoxicity (ADCC) mediated by 2H7scFv-IgG1 (SSS-S)H WCH2 WCH3:

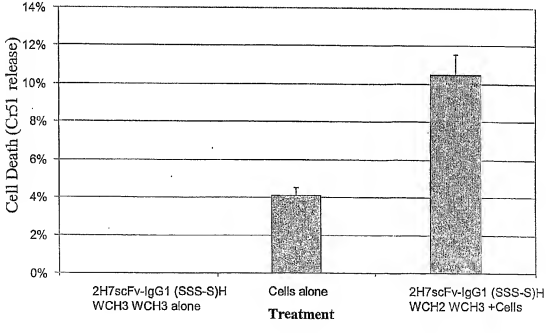


Fig. 5

Effects of Crosslinking of CD20 and CD40 Cell Surface Receptors on B Cell Proliferation:

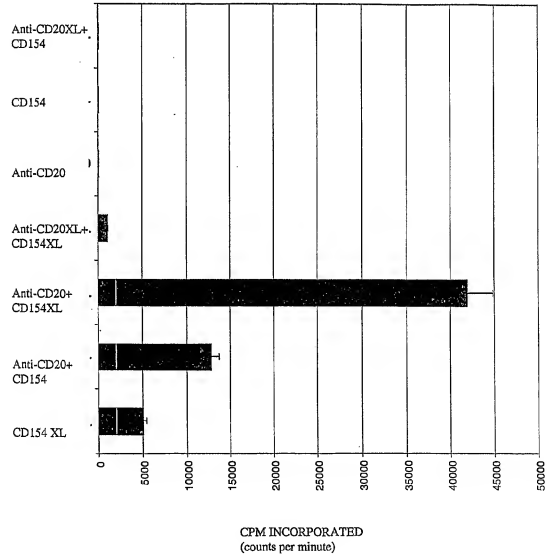


Fig. 6

Effect of Simultaneous ligation of CD20 and CD40 on CD95 and apoptosis.

Fig. 6A.

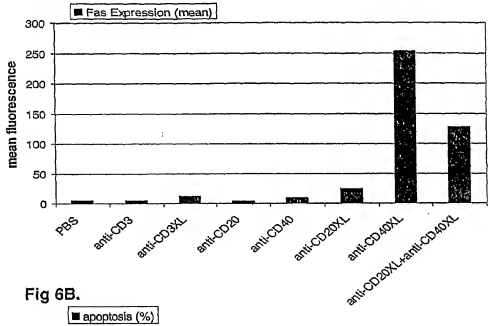


Fig 6B.

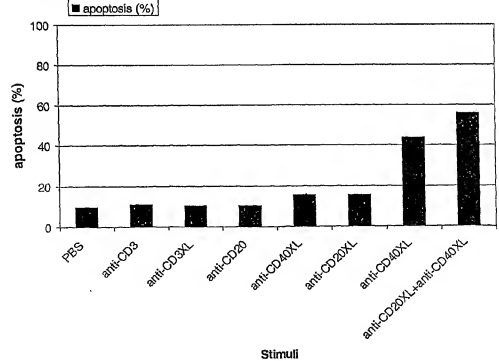


Fig. 7A

2H7-CD154 L2 cDNA and predicted amino acid sequence:

HindIII NcoI 2H7 V_L Leader Peptide →

```
                        M  D  F  Q  V  Q  I  F  S  F  L  L  I  S  A  S
1   AAGCTTGCCG CC   ATGGATTT TCAAGTGCAG ATTTTCAGCT TCCTGCTAAT CAGTGCTTCA

                                        2H7 V_L →
    V  I  I  A  R  G  Q  I  V  L  S  Q  S  P  A  I  L  S  A  S
61  GTCATAATTG CCAGAGGACA AATTGTTCTC TCCCAGTCTC CAGCAATCCT GTCTGCATCT

    P  G  E  K  V  T  M  T  C  R  A  S  S  S  V  S  Y  M  H  W
121 CCAGGGGAGA AGGTCACAAT GACTTGCAGG GCCAGCTCAA GTGTAAGTTA CATGCACTGG

                BamHI
    Y  Q  Q  K  P  G  S  S  P  K  P  W  I  Y  A  P  S  N  L  A
181 TACCAGCAGA AGCCAGGATC CTCCCCCAAA CCCTGGATTT ATGCCCCATC CAACCTGGCT

    S  G  V  P  A  R  F  S  G  S  G  S  G  T  S  Y  S  L  T  I
241 TCTGGAGTCC CTGCTCGCTT CAGTGGCAGT GGGTCTGGGA CCTCTTACTC TCTCACAATC

    S  R  V  E  A  E  D  A  A  T  Y  Y  C  Q  Q  W  S  F  N  P
301 AGCAGAGTGG AGGCTGAAGA TGCTGCCACT TATTACTGCC AGCAGTGGAG TTTTAACCCA

(Gly4Ser)3 Linker →
    P  T  F  G  A  G  T  K  L  E  L  K  G  G  G  G  S  G  G  G
361 CCCACGTTCG GTGCTGGGAC CAAGCTGGAG CTGAAAGGTG GCGGTGGCTC GGGCGGTGGT

                                                    2H7 V_H →
    G  S  G  G  G  G  S  S  Q  A  Y  L  Q  Q  S  G  A  E  L  V
421 GGATCTGGAG GAGGTGGGAG CTCTCAGGCT TATCTACAGC AGTCTGGGGC TGAGCTGGTG

    R  P  G  A  S  V  K  M  S  C  K  A  S  G  Y  T  F  T  S  Y
481 AGGCCTGGGG CCTCAGTGAA GATGTCCTGC AAGGCTTCTG GCTACACATT TACCAGTTAC

    N  M  H  W  V  K  Q  T  P  R  Q  G  L  E  W  I  G  A  I  Y
541 AATATGCACT GGGTAAAGCA GACACCTAGA CAGGGCCTGG AATGGATTGG AGCTATTTAT

    P  G  N  G  D  T  S  Y  N  Q  K  F  K  G  K  A  T  L  T  V
601 CCAGGAAATG GTGATACTTC CTACAATCAG AAGTTCAAGG GCAAGGCCAC ACTGACTGTA

    D  K  S  S  S  T  A  Y  M  Q  L  S  S  L  T  S  E  D  S  A
661 GACAAATCCT CCAGCACAGC CTACATGCAG CTCAGCAGCC TGACATCTGA AGACTCTGCG

    V  Y  F  C  A  R  V  V  Y  Y  S  N  S  Y  W  Y  F  D  V  W
721 GTCTATTTCT GTGCAAGAGT GGTGTACTAT AGTAACTCTT ACTGGTACTT CGATGTCTGG
```

Fig. 7A (continued)

human CD154/amino acid 48→

Bcl/Bam hybrid site

```
      G  T  G  T  T  V  T  V  S  D  P  R  R  L  D  K  I  E  D  E
781   GGCACAGGGA CCACGGTCAC CGTCTCTGAT CCAAGAAGGT TGGACAAGAT AGAAGATGAA

      R  N  L  H  E  D  F  V  F  M  K  T  I  Q  R  C  N  T  G  E
841   AGGAATCTTC ATGAAGATTT TGTATTCATG AAAACGATAC AGAGATGCAA CACAGGAGAA

      R  S  L  S  L  L  N  C  E  E  I  K  S  Q  F  E  G  F  V  K
901   AGATCCTTAT CCTTACTGAA CTGTGAGGAG ATTAAAAGCC AGTTTGAAGG CTTTGTGAAG

                                                          BclI
      D  I  M  L  N  K  E  E  T  K  K  E  N  S  F  E  M  Q  K  G
961   GATATAATGT TAAACAAAGA GGAGACGAAG AAAGAAAACA GCTTTGAAAT GCAAAAAGGT

      BclI
      ~~~~~
      D  Q  N  P  Q  I  A  A  H  V  I  S  E  A  S  S  K  T  T  S
1021  GATCAGAATC CTCAAATTGC GGCACATGTC ATAAGTGAGG CCAGCAGTAA AACAACATCT

      V  L  Q  W  A  E  K  G  Y  Y  T  M  S  N  N  L  V  T  L  E
1081  GTGTTACAGT GGGCTGAAAA AGGATACTAC ACCATGAGCA ACAACTTGGT AACCCTGGAA

      N  G  K  Q  L  T  V  K  R  Q  G  L  Y  Y  I  Y  A  Q  V  T
1141  AATGGGAAAC AGCTGACCGT TAAAAGACAA GGACTCTATT ATATCTATGC CCAAGTCACC

                  HindIII
                  ~~~~~~~
      F  C  S  N  R  E  A  S  S  Q  A  P  F  I  A  S  L  C  L  K
1201  TTCTGTTCCA ATCGGGAAGC TTCGAGTCAA GCTCCATTTA TAGCCAGCCT CTGCCTAAAG

      S  P  G  R  F  E  R  I  L  L  R  A  A  N  T  H  S  S  A  K
1261  TCCCCCGGTA GATTCGAGAG AATCTTACTC AGAGCTGCAA ATACCCACAG TTCCGCCAAA

      P  C  G  Q  Q  S  I  H  L  G  G  V  F  E  L  Q  P  G  A  S
1321  CCTTGCGGGC AACAATCCAT TCACTTGGGA GGAGTATTTG AATTGCAACC AGGTGCTTCG

                                          NcoI
                                          ~~~~~~
      V  F  V  N  V  T  D  P  S  Q  V  S  H  G  T  G  F  T  S  F
1381  GTGTTTGTCA ATGTGACTGA TCCAAGCCAA GTGAGCCATG GCACTGGCTT CACGTCCTTT

                  XhoI          XbaI
                  ~~~~~~        ~~~~
      G  L  L  K  L  E  *  *  S  R
1441  GGCTTACTCA AACTCGAGTG ATAATCTAGA
```

Fig. 7B.

2H7scFv-CD154 S4 cDNA and predicted amino acid sequence:

HindIII NcoI
~~~~~~ ~~~~~~2H7 V_L Leader Peptide→
M D F Q V Q I F S F L L I S A S
1 AAGCTTGCCG CC ATGGATTT TCAAGTGCAG ATTTTCAGCT TCCTGCTAAT CAGTGCTTCA

2H7 V_L →
V I I A R G Q I V L S Q S P A I L S A S
61 GTCATAATTG CCAGAGGACA AATTGTTCTC TCCCAGTCTC CAGCAATCCT GTCTGCATCT

P G E K V T M T C R A S S S V S Y M H W
121 CCAGGGGAGA AGGTCACAAT GACTTGCAGG GCCAGCTCAA GTGTAAGTTA CATGCACTGG

BamHI
~~~~~~~
Y Q Q K P G S S P K P W I Y A P S N L A
181 TACCAGCAGA AGCCAGGATC CTCCCCCAAA CCCTGGATTT ATGCCCCATC CAACCTGGCT

S G V P A R F S G S G S G T S Y S L T I
241 TCTGGAGTCC CTGCTCGCTT CAGTGGCAGT GGGTCTGGGA CCTCTTACTC TCTCACAATC

S R V E A E D A A T Y Y C Q Q W S F N P
301 AGCAGAGTGG AGGCTGAAGA TGCTGCCACT TATTACTGCC AGCAGTGGAG TTTTAACCCA

$(Gly_4Ser)_3$ Linker →
P T P G A G T K L E L K G G G G S G G G
361 CCCACGTTCG GTGCTGGGAC CAAGCTGGAG CTGAAAGGTG GCGGTGGCTC GGGCGGTGGT

2H7 V_H →
G S G G G G S S Q A Y L Q Q S G A E L V
421 GGATCTGGAG GAGGTGGGAG CTCTCAGGCT TATCTACAGC AGTCTGGGGC TGAGCTGGTG

R P G A S V K M S C K A S G Y T F T S Y
481 AGGCCTGGGG CCTCAGTGAA GATGTCCTGC AAGGCTTCTG GCTACACATT TACCAGTTAC

N M H W V K Q T P R Q G L E W I G A I Y
541 AATATGCACT GGGTAAAGCA GACACCTAGA CAGGGCCTGG AATGGATTGG AGCTATTTAT

P G N G D T S Y N Q K F K G K A T L T V
601 CCAGGAAATG GTGATACTTC CTACAATCAG AAGTTCAAGG GCAAGGCCAC ACTGACTGTA

D K S S S T A Y M Q L S S L T S E D S A
661 GACAAATCCT CCAGCACAGC CTACATGCAG CTCAGCAGCC TGACATCTGA AGACTCTGCG

V Y F C A R V V Y Y S N S Y W Y F D V W
721 GTCTATTTCT GTGCAAGAGT GGTGTACTAT AGTAACTCTT ACTGGTACTT CGATGTCTGG

Fig. 7B

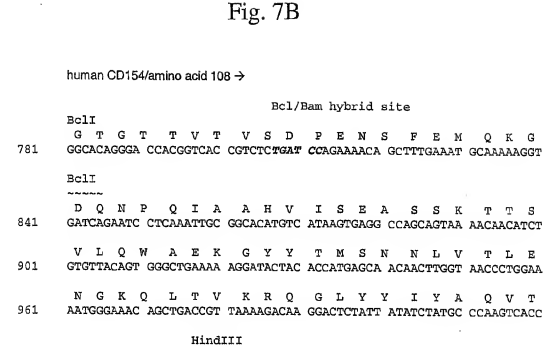
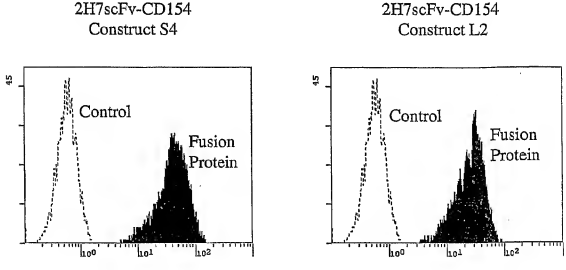


Fig. 8

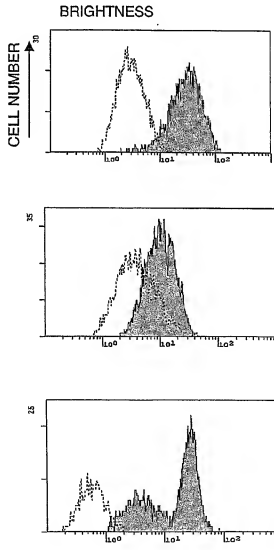
Simultaneous Binding of 2H7scFv-CD154 Fusion Proteins to CD20 and CD40



CD20 CHO cell targets + (control or fusion protein)
+ Biotin-CD40Ig + PE-SA

Fig. 9

Induction of Apoptosis Measured by Binding of Annexin V after incubation with 2H7scFv-CD154



.....control supernatant ___2H7scFv-CD154 supernatant

Fig. 10

Proliferation of T51 B Cell Line After Incubation with 2H7 scFv-CD154 S4 or 2H7 scFv-CD154 L2 Constructs

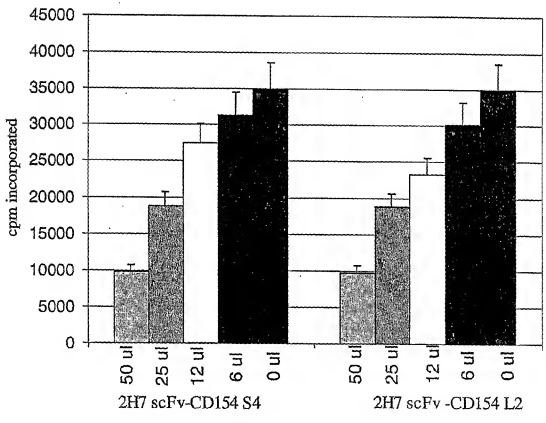


Fig. 11

Schematic Representation of 2H7 scFvIg Constructs

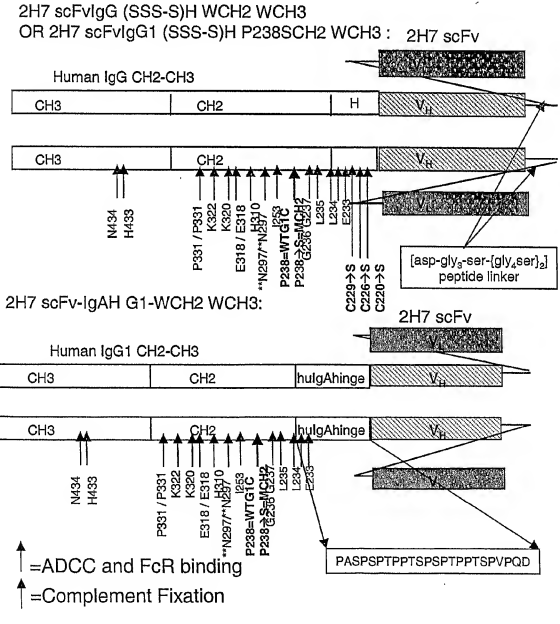


Fig. 12

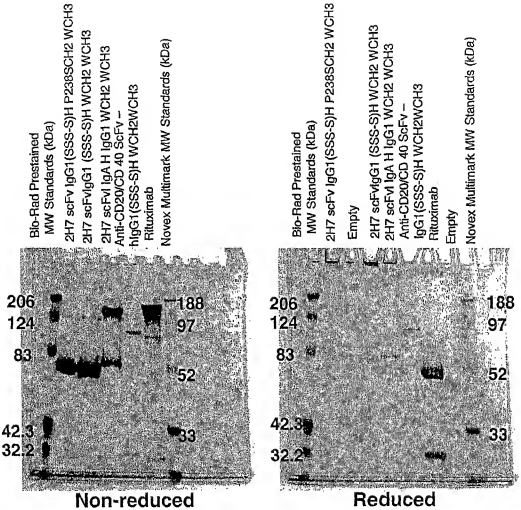


Figure 12: SDS-PAGE Analysis of CytoxB Derivatives. Purified fusion protein derivatives of CytoxB-scFvIg molecules and Rituximab were resuspended SDS sample buffer, boiled, loaded onto 10% Novex Tris-Bis gels (Invitrogen, San Diego, CA) and subjected to nonreducing (left panel) or reducing (right panel) SDS-PAGE electrophoresis at 175 volts. Two different molecular weight markers, BioRad prestained markers, and Novex Multimark molecular weight markers were also loaded onto each gel and the approximate size in kDa of each marker band is indicated along each side of the photographed gels. Gels were stained in Coomassie Blue stain and photographed with a SONY Mavica Digital camera. The mutant hinge forms of 2H7 scFvIgG1 migrate at approximately 70 kDa under both nonreducing and reducing conditions, indicating that these molecules are monomeric rather than dimeric in structure. The IgA hinge form of 2H7scFvIg migrates at approximately 75 kDa under reducing conditions, but migrates predominately as a dimer of 140 kDa with a fraction of the protein migrating at 75 kDa under nonreducing conditions. Under nonreducing conditions, rituximab migrates as a diffuse band of between 150 and 200 kDa. The heavy and light chains resolve into separate bands of approximately 32 and 50 kDa when rituximab is reduced and subjected to SDS-PAGE.

Fig. 13

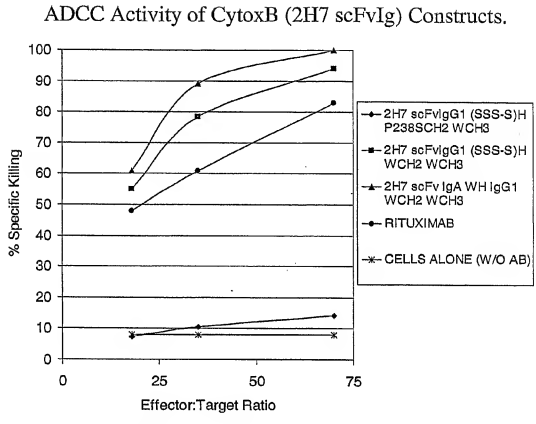


Figure 13: ADCC Activity of CytoxB Derivatives Compared to Rituximab. ADCC activity of CytoxB Derivatives or Rituximab was measured *in vitro* against BJAB B lymphoma cell line as target and using fresh human PBMC as effector cells. Effector to target ratios were varied as follows: 70:1, 35:1, and 18:1, with the number of BJAB cells per well remaining constant but varying the number of PBMC. Bjab cells were labeled for 2 hours with ^{51}Cr and aliquoted at a cell density of $5x10^4$ cells/well to each well of flat-bottom 96 well plates. Purified fusion proteins or rituximab were added at a concentration of 10 mg/ml, and PBMC were added at $9x10^5$ cells /well (18:1), $1.8x10^6$ cells/well (35:1), or $3.6x10^6$ cells/well (70:1), in a final volume of 200 µl. Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 4 hours, and 100 ml culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

Fig. 14

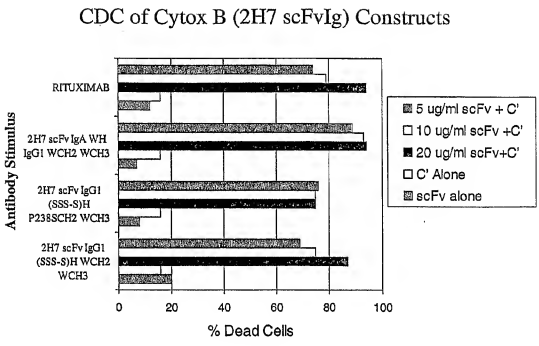
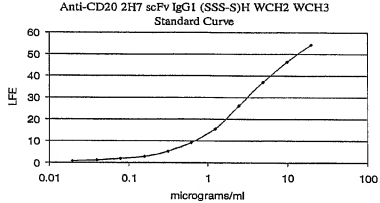


Figure 14: Complement Dependent Cytoxicity (CDC) Activity of CytoxB Derivatives Compared to Rituximab. 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3, 2H7 scFvIgG1 (SSS-S)H WCH2 WCH3, and 2H7scFv IgA WH IgG1 WCH2 WCH3 derivatives and Rituximab were compared for their ability to mediate complement dependent cytoxicity. Rabbit complement (Pel-Freez) was diluted 1:10 and added to BJAB cells along with dilutions of each antibody derivative (20 μg/ml, 10 μg/ml, and 5 μg/ml). Controls were also included without addition of complement (C') or scFv derivative. Reactions were allowed to continue for 1 hour, and cells from each well were then stained with trypan blue and the cell viability counted using a hemacytometer. Data is graphed as % of dead cells/total cells counted for each condition assayed.

Fig. 15

2H7 (anti-CD20) scFv IgG1 (SSS-S)H WCH2 WCH3 In Vivo Half Life



Macaque A99314

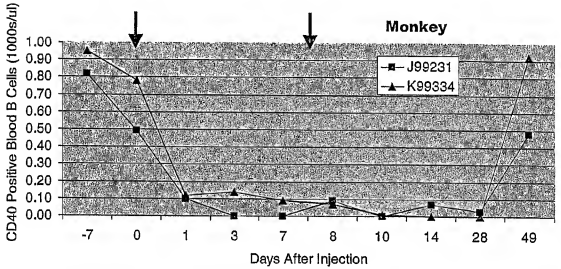
	Day	Binding intensity (LFE) @1:50 dilution of serum	estimated concentration (µg/ml)
	–7	0.213	<0.1
Injection #1 →	0	0.227	<0.1
	1	7.79	25.1
	3	5.51	15.6
Injection #2 →	7	3.37	9.4
	8	11.33	41.7
	10	5.45	15.4
	14	0.27	<0.1

Macaque F98081

	Day	Binding intensity (LFE) @ 1:50 dilution of serum	estimated concentration (µg/ml)
	–7	0.208	<0.1
Injection #1 →	0	0.219	<0.1
	1	6.73	21.9
	3	6.14	19.3
Injection #2 →	7	3.04	8.7
	8	9.83	33.8
	10	4.77	14.4
	14	0.231	<0.1

Fig. 16

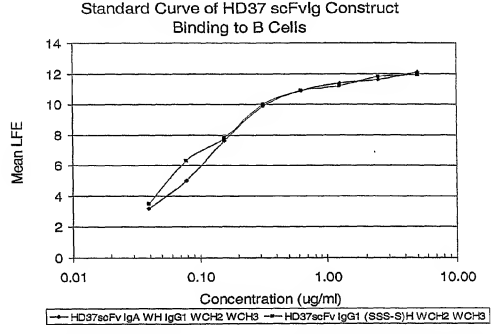
B Cell Depletion in macaques mediated by Cytox B20 (2H7 scFv IgG1 (SSS-S)H WCH2 WCH3) Construct



- **CytoxB20 injections of 6mg/kg yields 3 week B-cell depletion**
- **3-4 day half-life *in vivo***
- **CD20 saturation in lymph node B-cells at d14**
- **No first dose effects**
- **No anti-chimeric antibody development**

Fig. 17

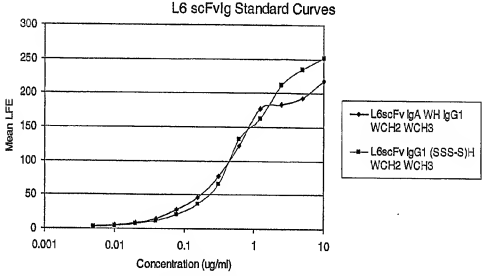
Production Levels of HD37 scFvIg Constructs by CHO Cell Lines



Clone/Isolate	Mean LFE at 1:100	Estimated Concentration
Bulk HD37 scFv IgA WH IgG1 WCH2 WCH3	11.2	> 60 ug/ml
1B2	10.4	>50 ug/ml
6C5	10.5	>50 ug/ml
4B1	8.6	>40 ug/ml
Bulk HD37 scFv IgG1 (SSS-S)H WCH2 WCH3	10.9	> 50 ug/ml
2G8	10.6	> 50 ug/ml
3F3	8.3	>40 ug/ml
3D9	11.1	> 60 ug/ml

Fig. 18

Production of L6 scFvIg constructs by CHO Cells



Construct	Mean LFE 1:20	Estimated Concentration
L6scFv IgA WH IgG1 WCH2 WCH3 unamplified CHO sup	51.1	6.25 ug/ml
L6scFv IgG1(SSS-S)H WCH2 WCH3 unamplified CHO sup	23.0	3.2 ug/ml

Fig. 19

ADCC Activity of 2H7 scFvIg, G28-1 scFvIg, and HD37 scFvIg Constructs

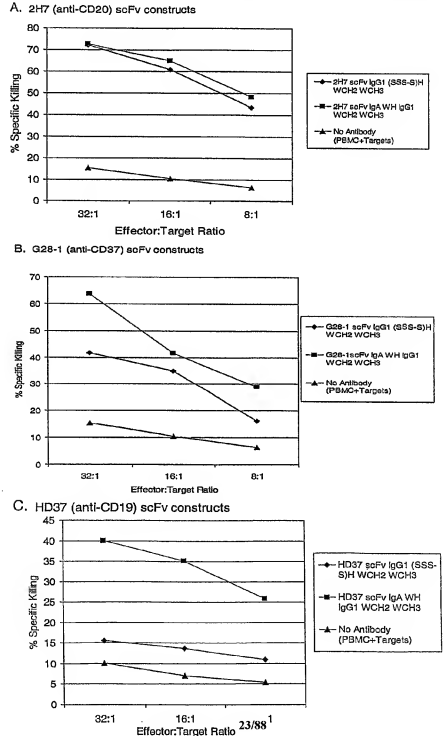


Fig. 20

ADCC Activity of L6 scFvIg Constructs

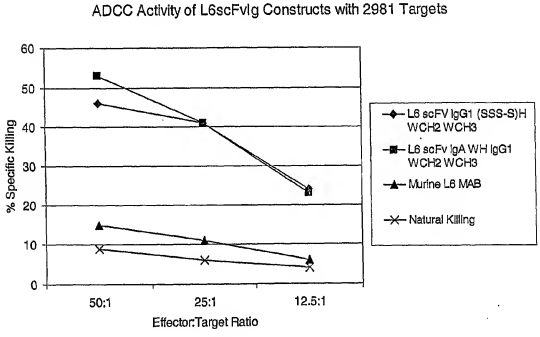


Fig. 21

SDS-PAGE Analysis of L6 and 2H7 scFvIg Fusion Proteins.

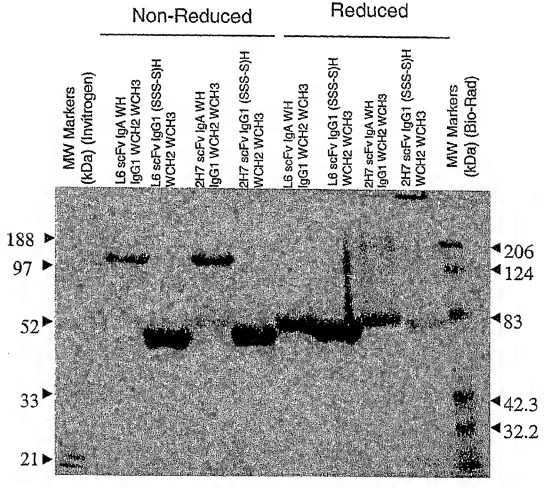


Fig. 22

SDS-PAGE Analysis of G28-1 and HD37 scFvIg Constructs.

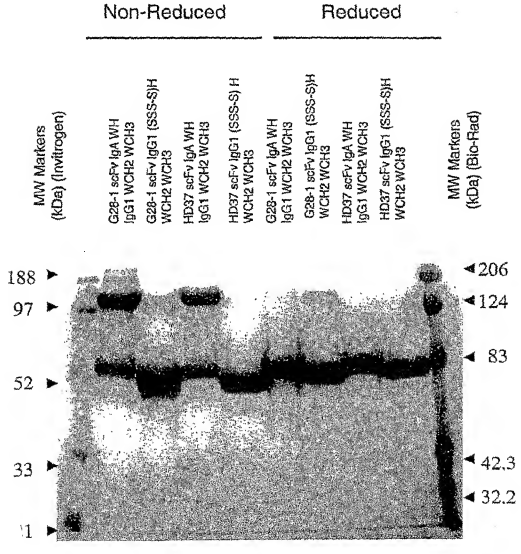


Fig. 23

Sequence alignment of human and llama Fc regions.

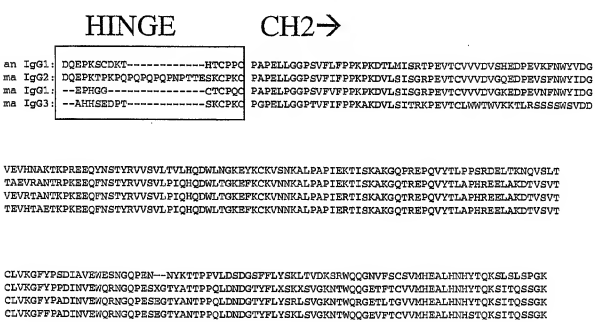


Fig. 24

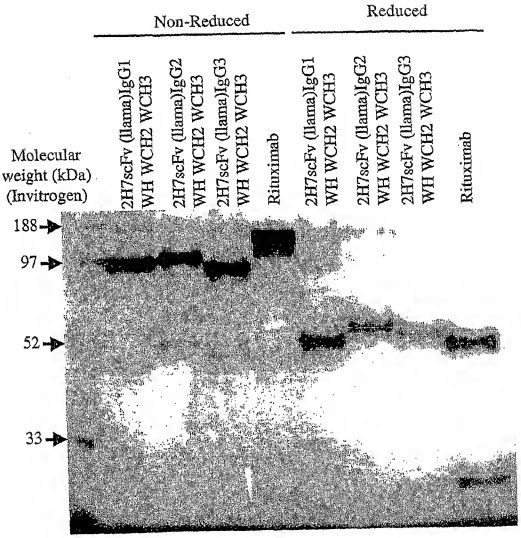


Fig. 25

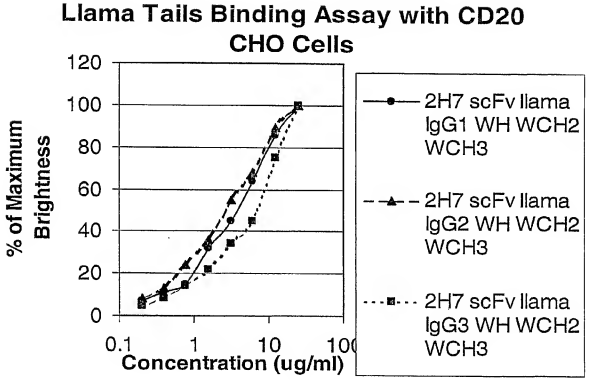


Fig. 26

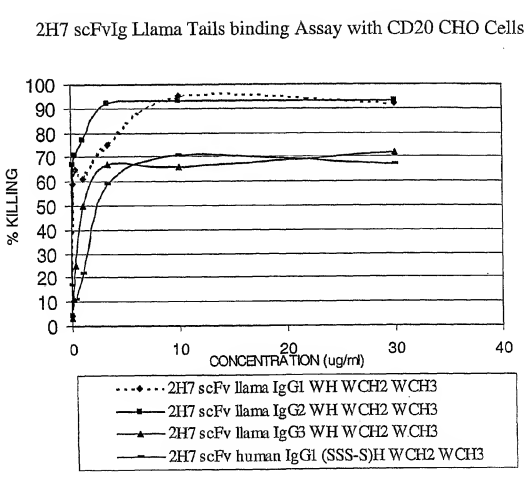


Fig. 27

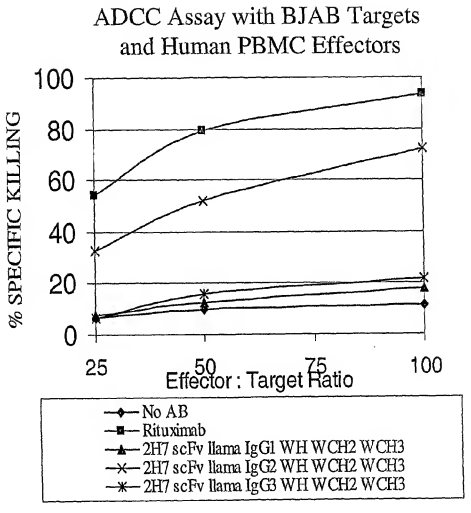


Fig. 28

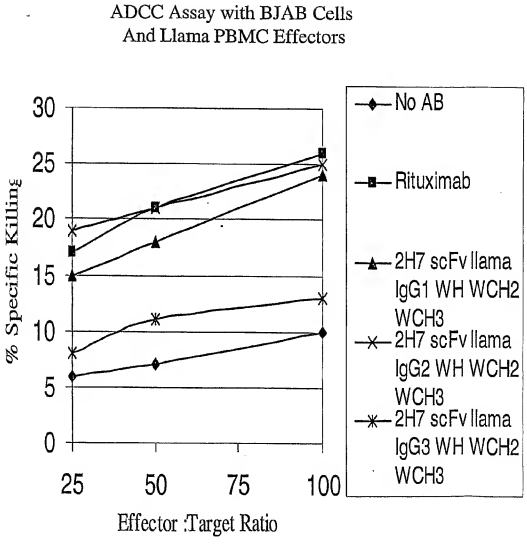
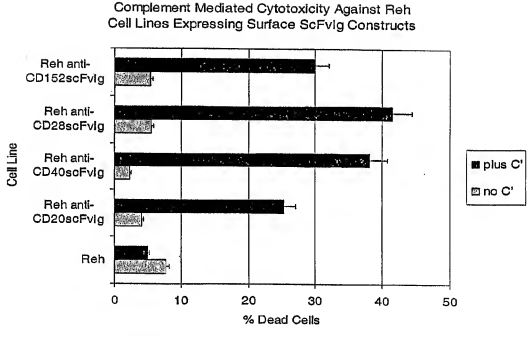
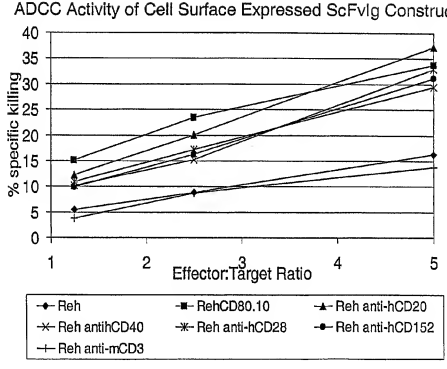


Fig. 29



Surface scFvIg and Effector Functions

Fig. 30



Surface scFvIg and Effector Functions

Fig. 31

Ig Constructs and Nomenclature:

Name Identifier	Hinge Sequence	CH2 Sequence	CH3 Sequence
hIgG1 (CCC-P)H WCH2 WCH3	IgG1 WT Hinge (CCC-P)	Wild Type CH2	Wild Type CH3
hIgG1 (SSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
VH L11S hIgG1 (SSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SSC-S)H WCH2 WCH3	IgG1 Mutant Hinge (SSC-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SCS-S)H WCH2 WCH3	IgG1 Mutant Hinge (SCS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (CSS-S)H WCH2 WCH3	IgG1 Mutant Hinge (CSS-S)	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgG1 (SSS-S)H P238S CH2 WCH3	IgG1 Mutant Hinge (SSS-S)	Mutant CH2 (IgG1) Pro→Ser 238	Wild type CH3 (IgG1)
IgA WH hIgG1 WCH2 WCH3	IgA Hinge	Wild type CH2 (IgG1)	Wild type CH3 (IgG1)
IgA WH IgA WCH2 WCH3	IgA Hinge	Wild type CH2 (IgA)	Wild type CH3 (IgA)
IgA WH IgA WCH2 T4CH3	IgA Hinge	Wild type CH2 (IgA)	Truncated CH3 (IgA) Missing 4 aa at COOH

Fig. 32

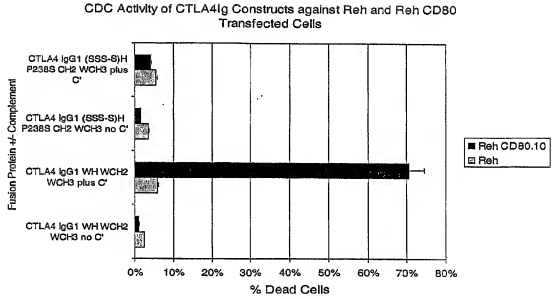


Fig. 33

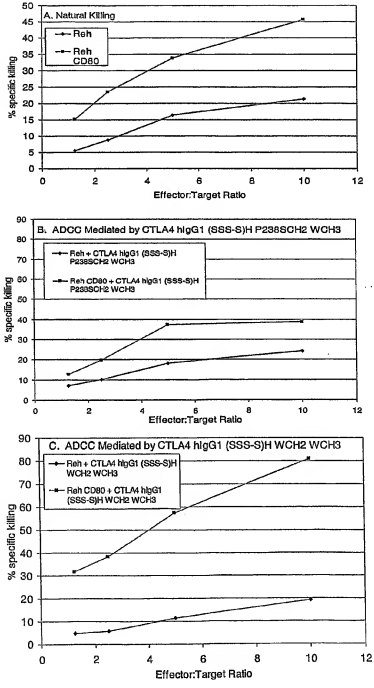


Fig. 34

Binding of 2H7 scFvIg Constructs
with Alternative Tails to CD20 CHO Cells

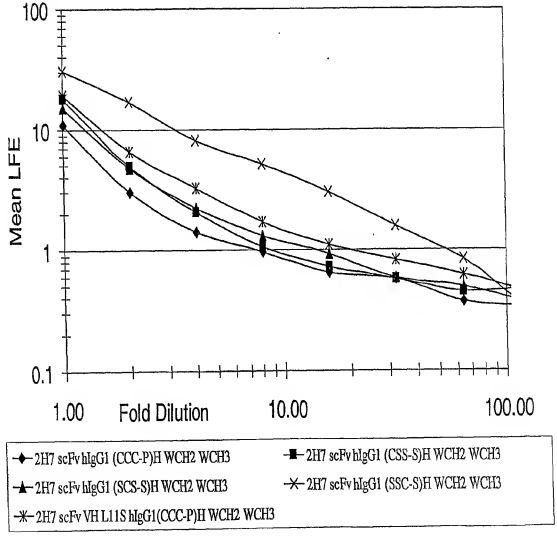


Fig. 35

Immunoblot Analysis of protein immunoprecipitates from COS transfections of 2H7 scFvIg Constructs

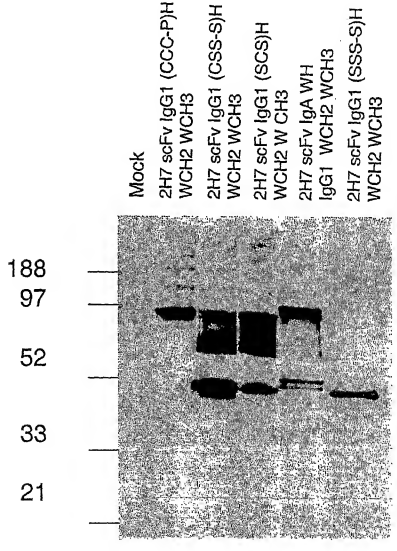


Fig. 36

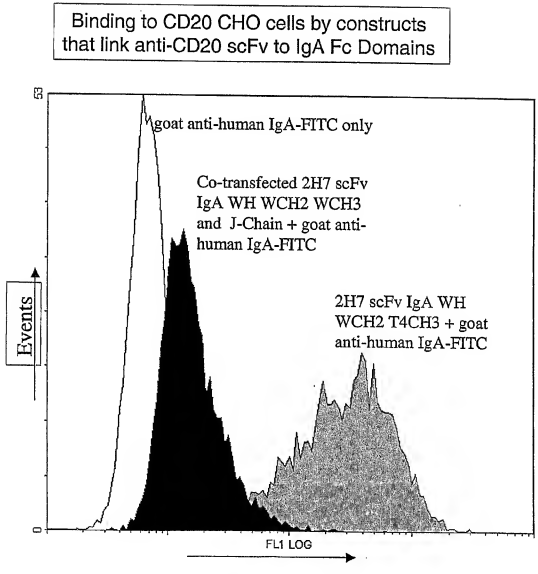


Fig. 37

Titration of CD20 specific scFvIg Constructs
for ADCC Activity Using Whole Blood Effectors

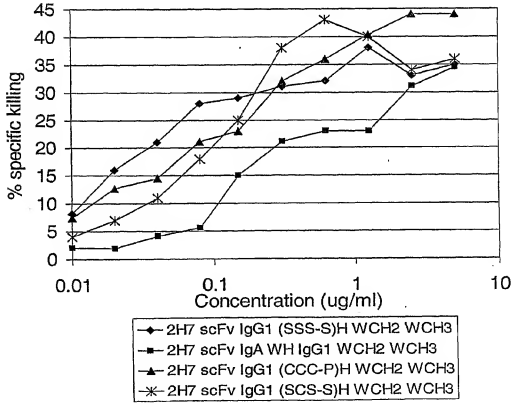


Fig. 38

ADCC Assay of anti-CD20 constructs with alternative tails
(Whole Blood Effectors / BJAB Targets

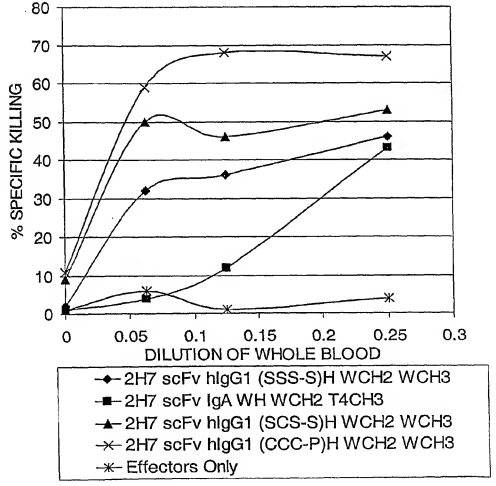


Fig. 39

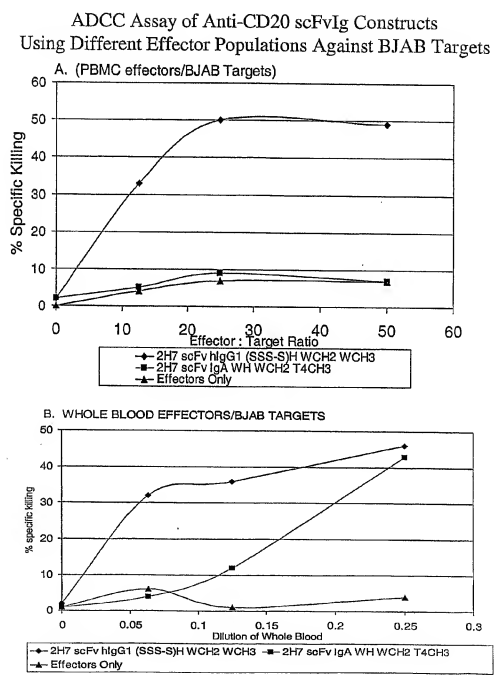
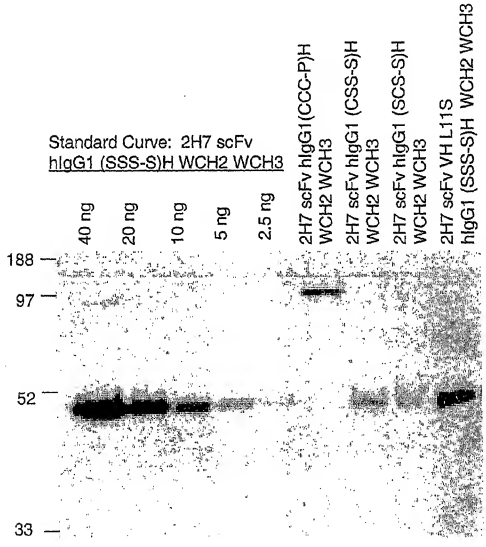


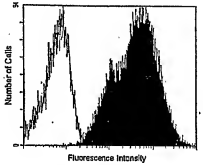
Fig. 40

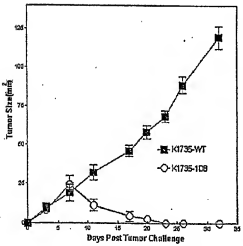
Immunoblot of 2H7 scFv Ig constructs from COS Transfections (1 µl/well) compared to a Concentration Standard

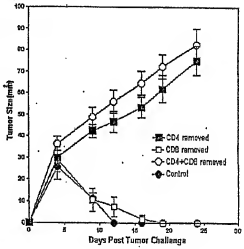


Figures 41A, 41B and 41C

A.







B.

C.

Fig. 42

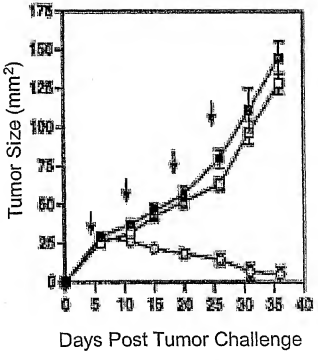


Fig. 43

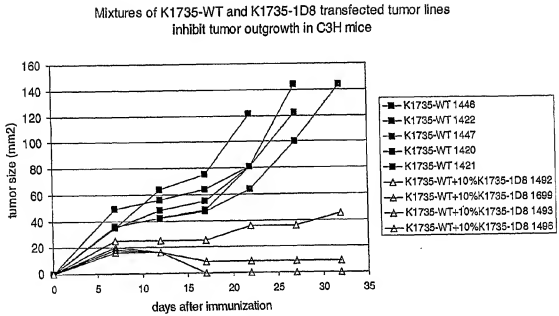


Fig. 44

Expression of anti-mouse CD137 (1D8) scFv-hIgG1 (SSS-S)H P238SCH2 WCH3
On the surface of panned Ag104-1D8 Transfected Tumor Cells

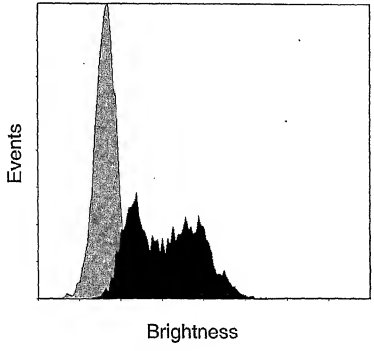


Fig. 45

Coomassie Stained SDS-PAGE Gel of 2H7 scFv Ig Constructs

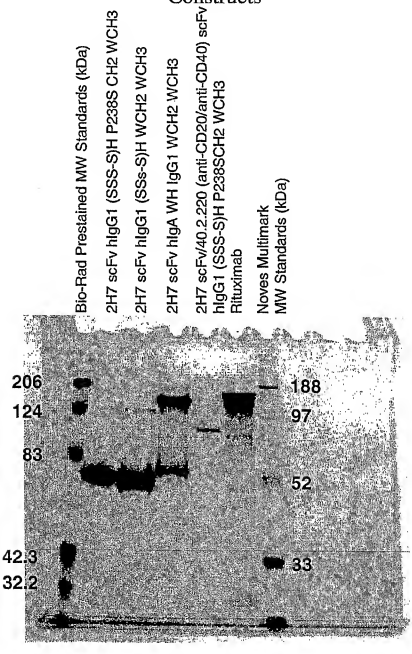


Fig. 46

ADCC mediated by 2H7 scFvIg Constructs by human PBMC effector cells against Bjab targets

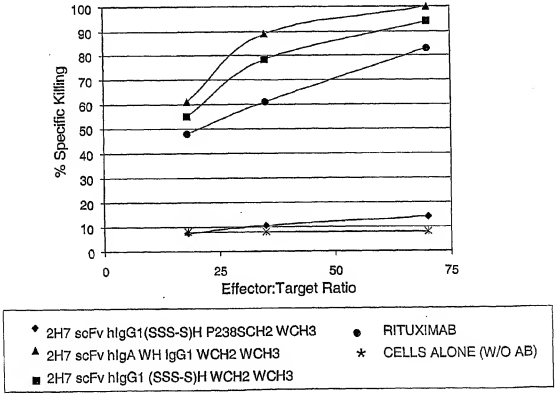
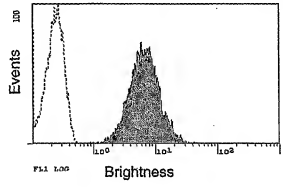


Fig. 47

Cell surface expression of anti-human CD3 G19-4 scFv hIgG1 (SSS-S)H P238SCH2 WCH3-hCD80TM/CT on Reh and T51 Cells.

Reh anti-CD3 (G19-4) scFv hIgG1 (SSS-S)H P238SCH2 WCH3-hCD80TM/CT



T51 G19-4 scFv hIgG1 (SSS-S)H P238SCH2 WCH3-hCD80TM/CT:

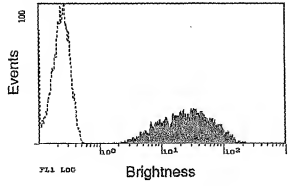
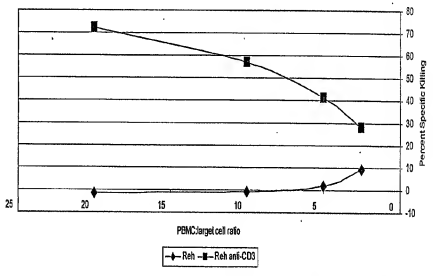


Figure 48.

Targeting of Cytotoxicity to Transfected Cell Lines by Surface expression of CD3 scFvIg

A



B

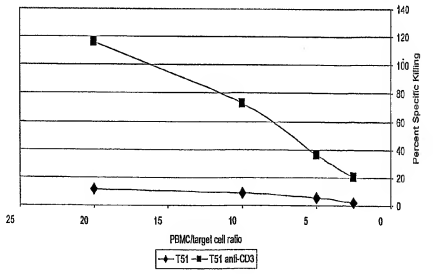


Fig. 49

Binding of 5B9, a mouse anti-human CD137 scFv hIgG1 (SSS-S)H WCH2WCH3 to stimulated human PBMC

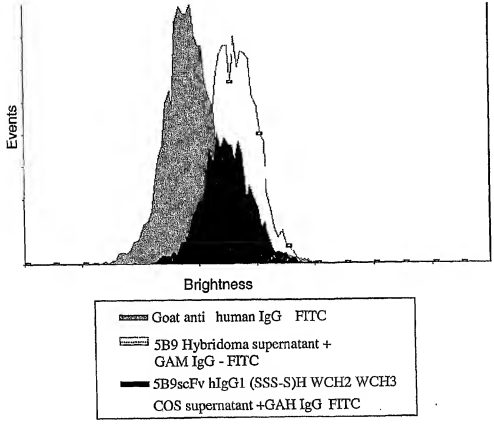
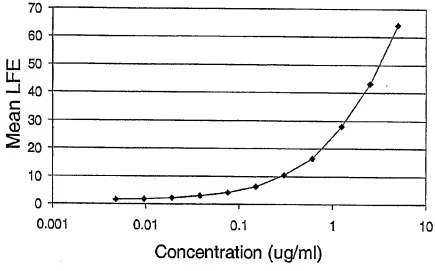


Fig. 50

Effect of V_HL11S Mutation on CytoxB20
2H7 scFv hIgG1 (SSS-S)H WCH2 WCH3 Protein Expression

50A. Standard Curve: 2H7VH-L11S-IgG1 (SSS-S)H WCH2 WCH3



50B. CHO supernatant Brightness and Estimation of Protein concentrations from Standard Curve:

	CHO clone name				
	4F2	**4F5**	**3E5**	**6B11A**	**2B8A**
Mean LFE					
1/100	71.7	40.6	31.5	99.7	101.5
1/500	27.1	12.4	11.2	40.8	43
approx conc. µg/ml	**600**	**225**	**125**	**1000**	**1250**

Fig. 51

Production Levels of 2H7scFv VH L11S hIgG1 (SSS-S)H WCH2 WCH3 From CHO Clone Culture Supernatants

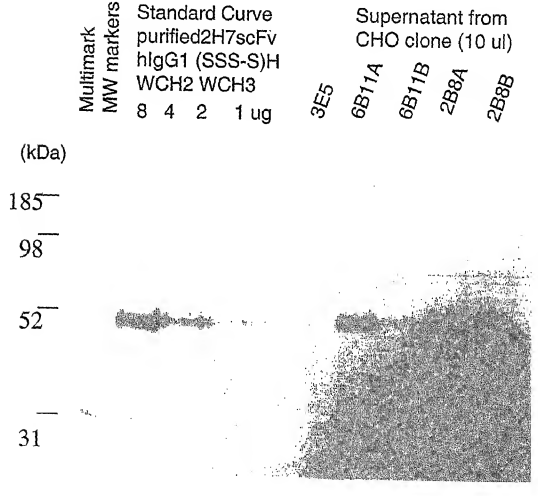


Fig. 52

Effect of VHL11S Mutation on G28-1 scFvIg Construct Protein Production from COS cells

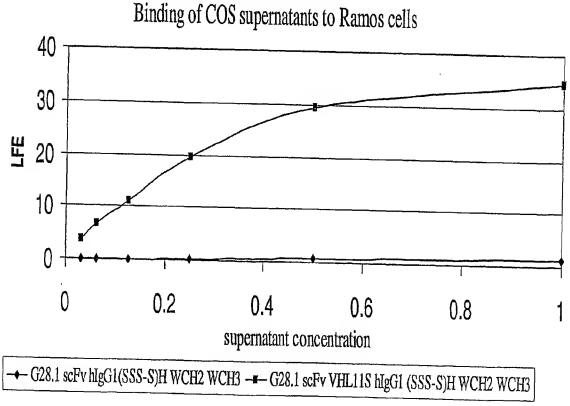


Fig. 53

Immunoblot of G28-1 scFvIg Constructs

Increased Protein Levels in COS supernatants
transfected with G28-1scFv hIgG1 (SSS-S)H WCH2 WCH3
After Substitution of Leucine with Serine at position 11 of VH (VHL11S)

Fig. 53A.

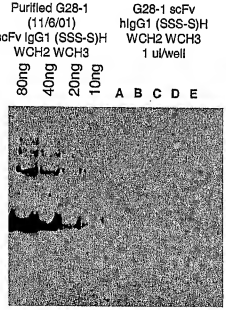


Fig. 53B.

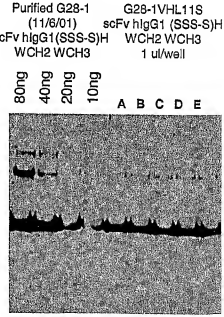


Fig. 54

Binding of 2H7 scFvIg Constructs with Altered Hinges and CH3 domains to CD20 CHO Cells

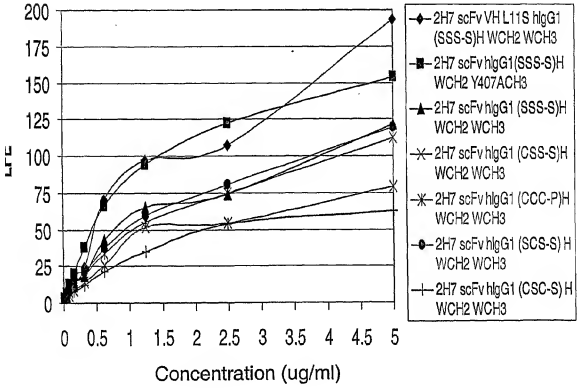


Fig. 55

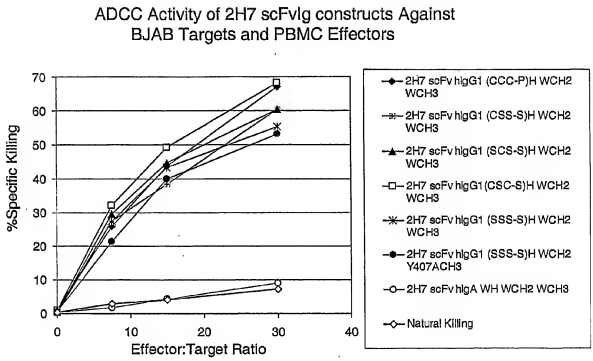


Fig. 56

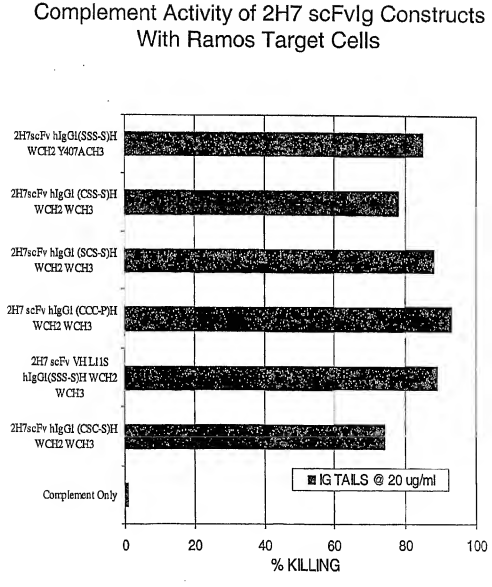
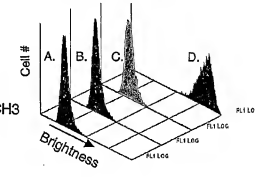


Fig. 57

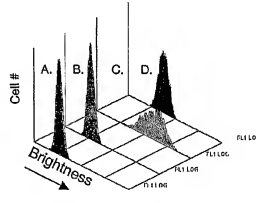
Binding of 2H7 scFvIg Derivatives to CD20CHO Cells

A. No fusion protein
B. 2H7 scFv hIgE CH2CH3CH4
C. 2H7 scFv hIgA WH WCH2 WCH3
D. 2H7 scFv hIgG1 (SSS-S)H WCH2 WCH3

FITC anti-hIgG



FITC anti-hIgA



FITC anti-hIgE

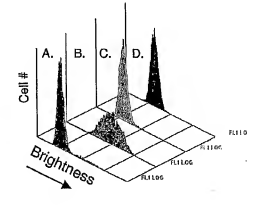


Fig. 58

Fig. 58A. 2H7 scFv VH L11S human IgE (WCH2 WCH3 WCH4) Binding to CD20 CHO at 30 ug/ml

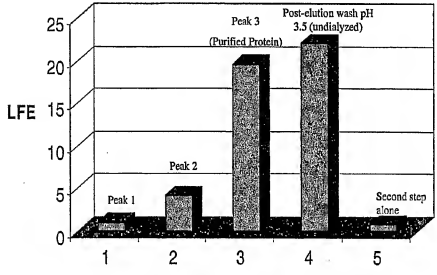


Fig. 58B. ADCC Activity of 2H7 VHL11S IgE (WCH2 WCH3 WCH4) Protein Fractions with **PBMC** Effectors and Bjab Targets

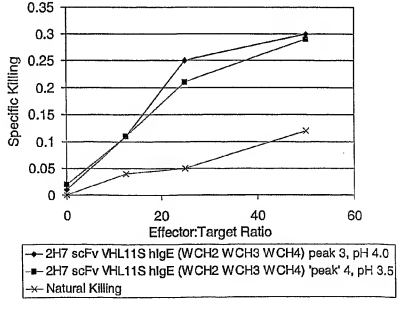


Fig. 59

Binding Data for COS derived α-CD20 (2H7) scFv VHL11S mIg E (WCH2 WCH3 WCH4) and mIgA (WH WCH2 WCH3)Tailed Molecules

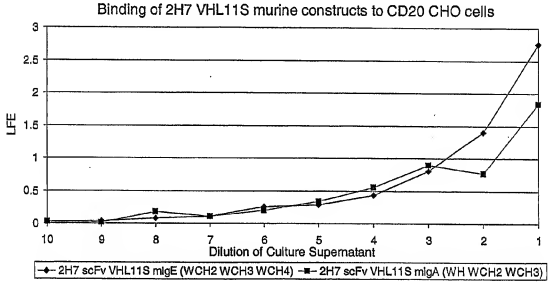


Fig. 60

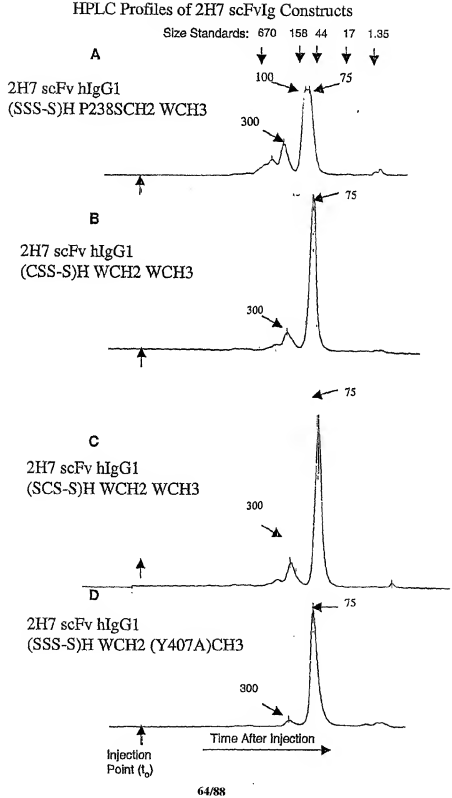


Fig. 61

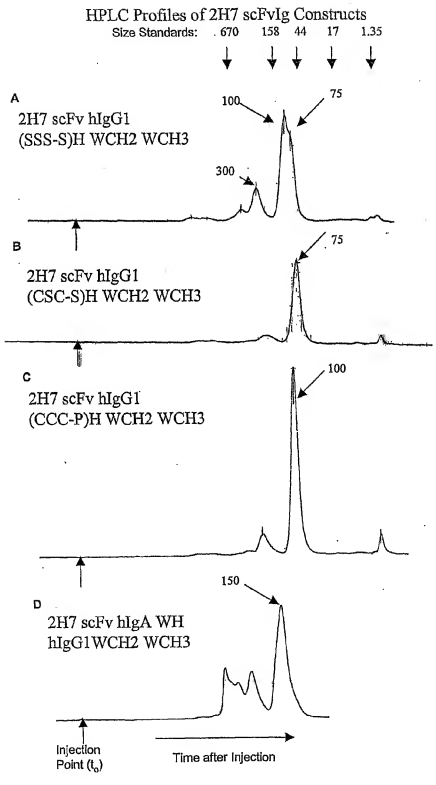


Fig. 62

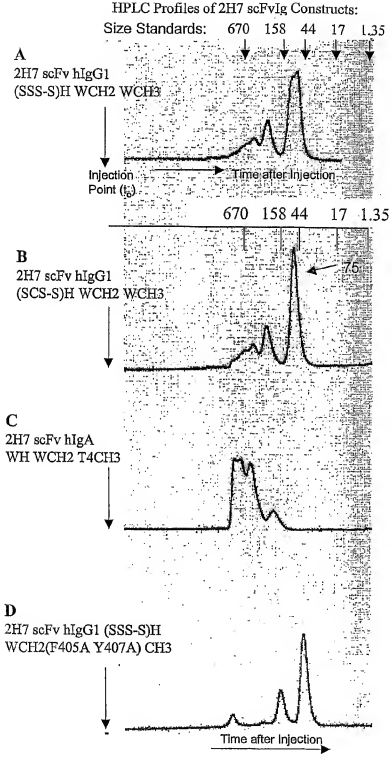


Fig. 63

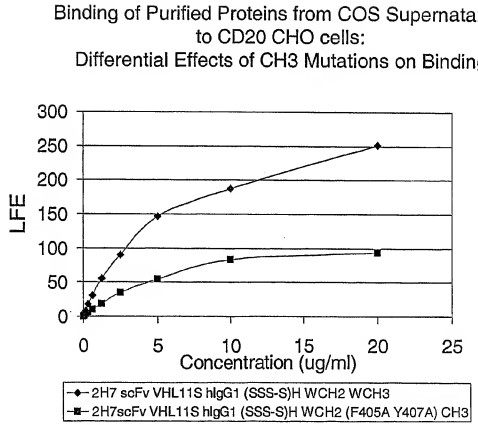


Fig. 64

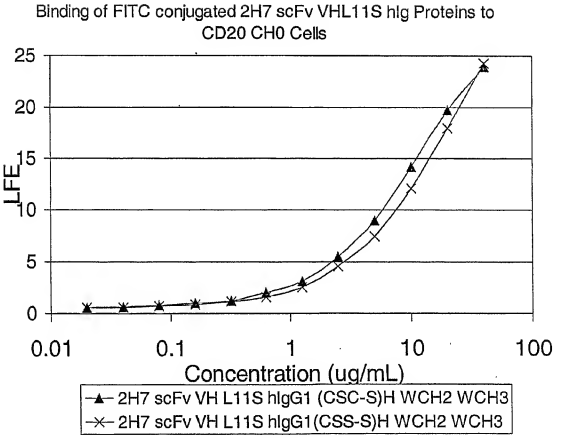


Fig. 65

Nonreducing SDS-PAGE on Protein A-Purified Lots of 2H7 scFv VHL11S hIg Constructs (10 ug/lane)

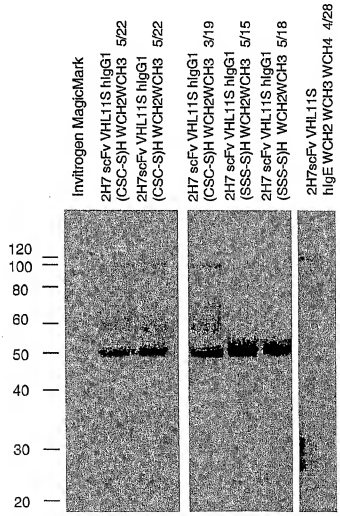


Fig. 66

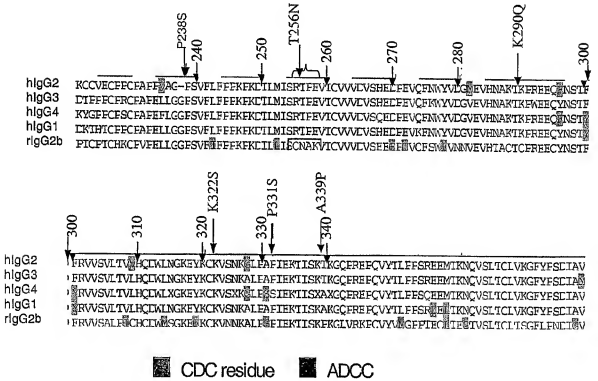


Figure 67.

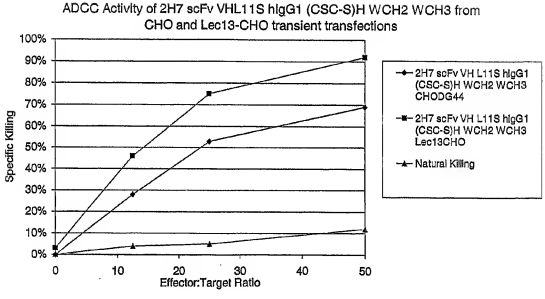


Fig. 68

CD16(ED) hIgG1(SSS-S)H P238S CH2 WCH3 high and low affinity alleles expressed as soluble molecules

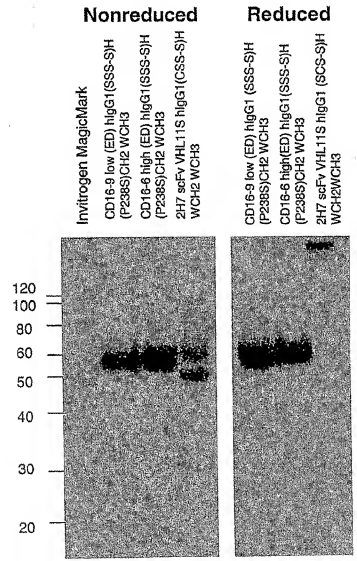


Fig. 69

Binding of soluble CD16-FITC high and low affinity fusion proteins to 2H7 scFv VHL11S hIgG1 (CSC-S)H WCH2WCH3 or (SSS-S)H (P238S)CH2WCH3 on CD20CHO Targets

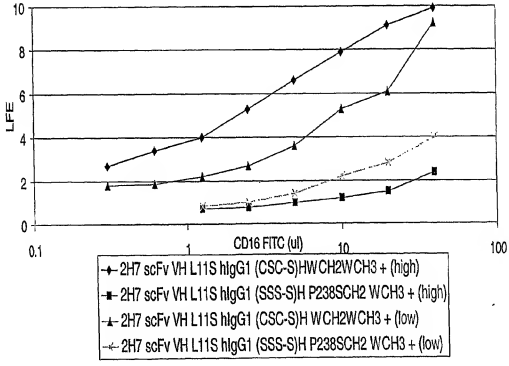
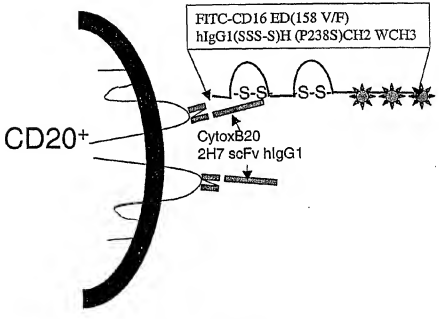


Fig. 70

Binding of FITC Labeled, Recombinant Human CD16(ED) extracellular domain -Ig Fusion Protein to CytoxB Derivatives on CD20 CHO Cells



Expression of surface displayed SMIPs links modified cDNAs with the altered fusion proteins

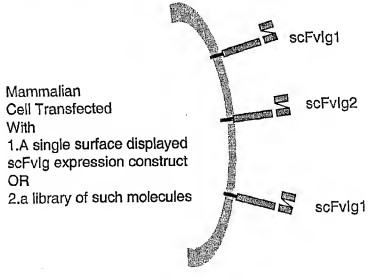


Fig. 71

CD37 mAbs and scFvIg Induce Apoptosis

	Bjab Staining	Annexin V Positive	
	No scFvIg	17.5	
	2H7 MH	27	
scFvIg	G28-1 MH	30.6	
	G28-1 IgAH	28.9	
	HD37 MH	29.1	
	(2H7+G28-1)MH	41	
	(2H7+HD37) MH	37.1	
	(G28-1+HD37) MH	35.3	
			plus GAM
	Ramos	AnnexinV Positive	AnnexinV positive
	cells alone	3	3.3
	2H7 Mab	1.4	3.1
	G28-1 Mab	18.3	8.7
mAbs	HD37 Mab	3.7	3.1
	G28-5	3.9	8.3
	2H7+G28-1	32.3	35.7
	2H7+HD37	5	10.5
	2H7+G28-5	5.7	19.4
	HD37+G28-1	26.9	50
	HD37+G28-5	8.2	18.4
	G28-1+G28-5	39.5	68.3

Caspase 3 Activity in Ramos Cells after 4 Hour Incubation With CytoxB20G SMIP

Fig. 72

Fig. 73

Figure 76: CDC Activity of CytoxB20G SMIPS. CytoxB20G, CytoxB20GP238, or Rturximab were incubated at increasing concentrations with 10^4 Bjab Target Cells and a 1:10 dilution of rabbit complement (Pelfreez) in a volume of 100 microliters for sixty minutes. Aliquots were stained with trypan blue (Invitrogen), and counted using a hemacytometer to determine the percentage of the cell population killed during treatment. Negative controls with cells and only one reagent were also included.

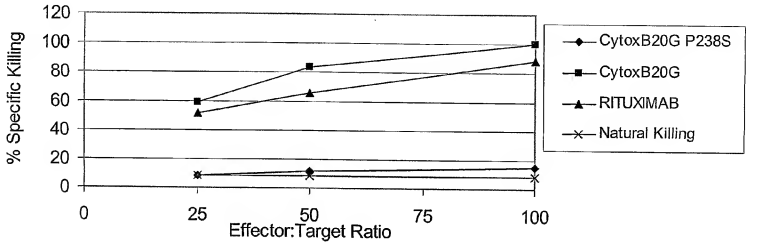


Fig. 74

Figure 77: ADCC Activity of CytoxB20G SMIPS. ADCC activity of CytoxB20G or Rituximab was measured *in vitro* against BJAB B lymphoma cell line as target and using fresh human PBMC as effector cells. Effector to target ratios were varied as follows: 100:1, 50:1, and 25:1, with the number of BJAB cells per well remaining constant but varying the number of PBMC. Bjab cells were labeled for 2 hours with ^{51}Cr and aliquoted at a cell density of $5x10^4$ cells/well to each well of flat-bottom 96 well plates. Purified fusion proteins or rituximab were added at a concentration of 10 μg/ml, and PBMC were added at 1.25 x 10^6 cells /well (25:1), 2.5 x 10^5 cells/well (50:1), or 5 x 10^6 cells/well (100:1), in a final volume of 200 μl. Natural Killing was measured at each effector:target ratio by omission of SMIP or MAb. Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 5 hours, and 100 μl culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

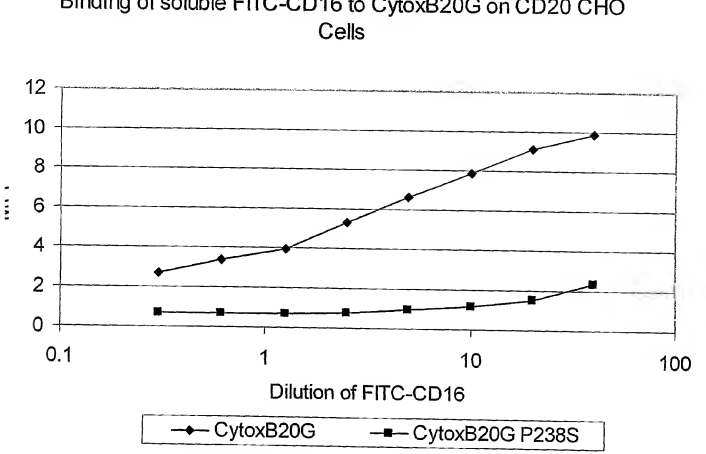


Fig. 75

re 78: Binding of soluble FITC-CD16 to CytoxB20G on CD20 CHO cells. CD20 CHO cells (10^6) were incubated
ı saturating amounts of CytoxB20G or CytoxB20G P238S(10 ug/ml) for one hour on ice in PBS/2% FBS. Cells
: washed in PBS/2% FBS and incubated with serial dilutions of 0.5 mg/ml FITC-CD16 for one hour on ice. Cells
: washed and specific binding measured by flow cytometry using a Beckman-Coulter Epics C machine. Results
: analyzed using Expo analysis software and normalized fluorescence units graphed as a function of concentration.

Fig. 76

re 79: CytoxB20G SMIPs bind similarly to U937 cells expressing the high affinity FcR (FcγRI, CD64). U937 cells
expressing CD64 were incubated in PBS/2%FBS for one hour on ice with CytoxB20G orCytoxB20G P238S. Cells
washed and incubated for one hour on ice with FITC-goat anti-human IgG1 (Fc specific) (Caltag) at a final dilution
100. Cells were washed and fluorescence analysed on a Beckman-Coulter EpicsC flow cytometer. Data was
yzed using Expo analysis software, and fluorescence intensity graphed as a function of SMIP concentration.

Fig. 77

10: CytoxB20G or CytoxB20G P238S were administered to macaques by intravenous injection at 6 mg/kg, with isions given one week apart. The effect on circulating B cells was measured by detection of CD40 positive B cells neral blood. Blood samples were drawn from injected animals at days -7,0,1,3,7,8,10,14, 28, and 43. B cell n was estimated by performing CBC (complete blood counts) and two color flow cytometry analysis on monkey FITC or PE conjugates of antibodies against CD40, CD19,CD20, IgG, CD3, CD8 were used in various tions. Data are plotted as the number of CD40 positive blood B cells tabulated in thousands of cells per er over time relative to the initial pre-injection time point level of B cells (maximum).

Figure 81: SEC on CytoxB37G SMIPs containing SSS and SSC hinge Domains from Human IgG1

Fig. 78

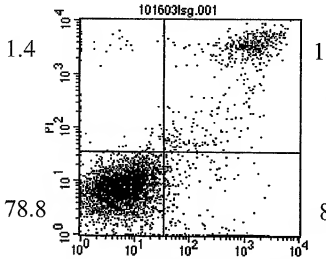
Figure 81: SEC (Size Exclusion Chromatography) CytoxB37G SMIPs were purified from CHO culture supernatants by Protein A affinity chromatography. Purified aliquots of 10-25 µg were subjected to HPLC over a Tosoh Biosep, Inc. TSK 3000 SWXL HPLC column, poi size 5 µm. The flow rate was 1ml/min, in PBS, pH 7.2 running buffer. Migration rates of molecular weight standards are indicated below the tracing. The CytoxB37G (SSS)H SMIP indicated in blue, while the CytoxB37G (CSS)H is indicated in red.

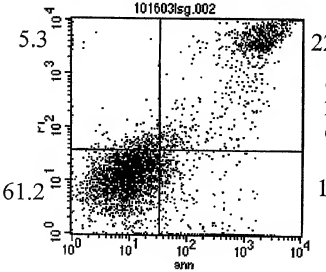
gure 82: Binding of CytoxB37G SMIPs to B Cell Lymphoma Cell Lines

Fig. 79

ure 82 : Binding of CytoxB37G SMIPS to B cell lymphoma cell lines. Serial dilutions of ified CytoxB37 (SSS)H G or CytoxB37 (SSC)H G SMIPS were incubated with 10^6 cells of each cell type for 60 utes on ice in PBS/2%FBS. Samples were washed twice, and incubated with a mixture of FITC goat anti-human i and FITC goat anti-human IgG F(ab')2 (CalTag) at 1:100 each, on ice for 45 minutes. Samples were washed and lyzed by flow cytometry using a FACsCalibur (Becton-Dickinson)

Fig. 80: AnnexinV-PI Staining of Ramos Cells Incubated 24 hours with CD37 SMIPS





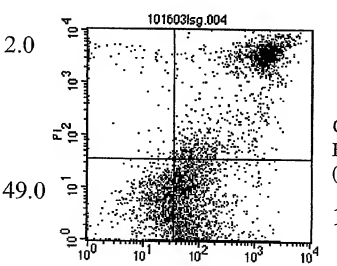


Figure 84: Thymidine Incorporation (Growth Inhibition) in Ramos B-cells after a 48 Hour Incubation with anti-CD37 SMIPS

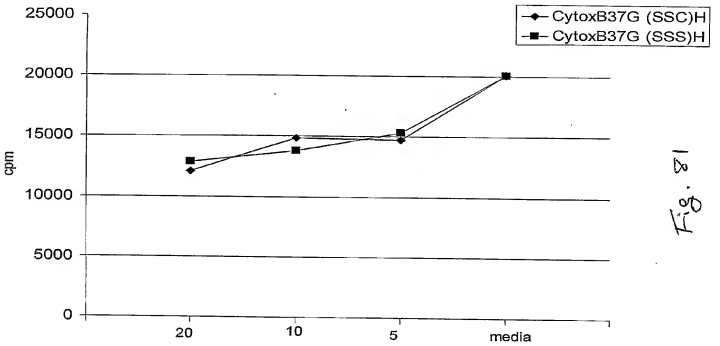


Fig. 81

ure 84: Ramos B cells were incubated with serial dilutions of purified CD37G SMIPS containing er the IgG1 hinge identified as (SSS)H or (SSC)H. Cultures were incubated in 96 well flat bottom ie culture dishes (Costar) at 37°C, 5%CO_2 for 36 hours prior to pulsing with ^{3}H-thymidine for the 12 hours of a 48 hour incubation (0.75 µCi/well). Cells were harvested onto 96-well GFC plates g a Packard harvester, dried, and 25 µl Microscint scintillation fluid added to each well prior to nting on a TopCount NXT microplate (Packard) scintillation counter. Data are plotted as cpm rporated versus protein concentration. Each SMIP show increasing inhibition of proliferation with easing protein concentration.

ure 85: The Induction of Apoptosis in Ramos B-cells after a 20 hour incubation with different combinations of CD20 and CD37 targeted SMIPS

Fig. 82

gure 85: Ramos B cells were incubated with CD20 and/or CD37 targeted SMIPs (10 /ml) in solution for 20 hours. Cells were then harvested, washed, and incubated in nexinV and propidium iodide using a staining kit from Immunotech prior to two color w cytometry using a FACsCalibur flow cytometer (Becton-Dickinson). The graph show › percentage of annexin V positive cells identified by their staining in the right quadrants the dot plots.

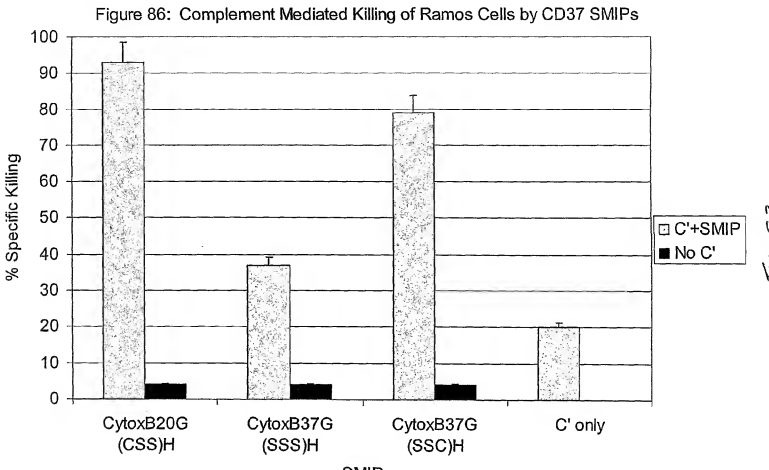


Fig. 83

Figure 86: CDC Activity of CytoxB37G SMIPS. CytoxB20G, CytoxB37 (SSS)H G, CytoxB37 (SCS)H, CytoxB37 (CSS)H, or CytoxB37 (SSC)H were incubated at 10 μg/ml with 10^4 Ramos Target Cells and a 1:10 dilution of rabbit complement (PelFreez) in a volume of 150 μl for 90 minutes. Aliquots were stained with trypan blue (Invitrogen), and counted using a hemacytometer to determine the percentage of the cell population killed during treatment. Negative controls with cells and only one reagent were also included.

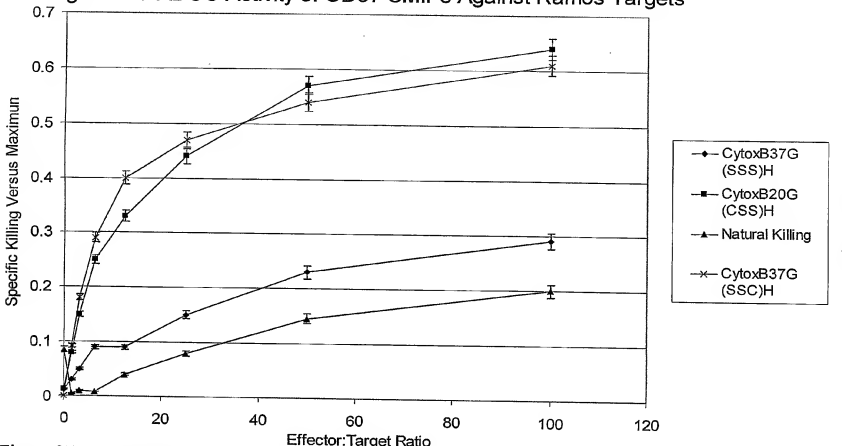


Fig. 84

Figure 87: CD37 SMIPS at 10 μg/ml were incubated in flat-bottom 96 well plates with 10^4 ^{51}Cr-labeled Ramo cells and resting human PBMCs at different effector:target ratios ranging from 0 to 100. All incubations were performed in triplicate at each effector:target ratio. Natural Killing was measured at each effector:target ratio by omission of SMIP. Spontaneous release was measured without addition of PBMC or fusion protein, and maximal release was measured by the addition of detergent (1% NP-40) to the appropriate wells. Reactions were incubated for 6 hours, and 100 ml culture supernatant harvested to a Lumaplate (Packard Instruments) and allowed to dry overnight prior to counting cpm released on a Packard Top Count NXT Microplate Scintillation Counter.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US03/41600

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C12N 15/00; A61K 39/395; C07K 16/00
US CL : 530/387.3, 388.85, 391.3; 424/130.1; 536/23.4; 435/320.1, 69.6

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
U.S. : 530/387.3, 388.85, 391.3; 424/130.1; 536/23.4; 435/320.1, 69.6

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Continuation Sheet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	HAYDEN et al. Single-chain mono- and bispecific antibody derivatives with novel biological properties and antitumor activity from COS cell transient expression system. Therapeutic Immunology. 1994, Vol. 94, pages 3-15, especially Figure 1, Methods.	1-7, 20-28, 31-40, 53-57, 59, 62-63, 65-75, 116-119, 129-137, 140-150, 161-169, 171-181, 238, 240-243, 251-259, 261-267, 282-285, 287-295, 399-411
Y	US 6,147,203 A (PASTAN et al.) 14 November 200 (14.11.2000), see entire document, especially abstract, column5-6.	1-7, 20-28, 31-40, 53-57, 59, 62-63, 65-75, 116-119, 129-137, 140-150, 161-169, 171-181, 238, 240-243, 251-259, 261-267, 282-285, 287-295, 399-411

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

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"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
29 October 2004 (29.10.2004)	02 NOV 2004
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US Commissioner for Patents P.O. Box 1450 Alexandria, Virginia 22313-1450 Facsimile No. (703) 305-3230	Authorized officer Larry R. Helms Telephone No. 571-272-1600

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INTERNATIONAL SEARCH REPORT

PCT/US03/41600

C. (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 6,074,644 A (PASTAN et al.) 13 June 2000 (13.06.2000), see entire document, especially column 20.	26, 28, 32, 135, 137, 142, 257, 259, 262
Y	US 5,677,425 A (BODMER et al.) 14 October 1997 (14.10.1997), see entire document, especially abstract, column 3-4.	65-75, 116, 172-180, 288-295
Y	US 6,482,919 B2 (LEDBETTER et al.) 19 November 2002 (19.11.2002), see entire document.	180

INTERNATIONAL SEARCH REPORT

PCT/US03/41600

Continuation of B. FIELDS SEARCHED Item 3:
CAPLUS, MEDLINE, WEST, BIOSIS
Search terms: inventor name, scfv, hinge, cysteine, fusion protein, CD19, CD3, deleted hinge, altered hinge, IgG1, IgA, IgE, disulfide stabilized, constnat region.